

When treated with a Src inhibitor (CGP77675, 1 μ M) added 24h prior to and at 24h intervals during the course of oxaliplatin-exposure, oxaliplatin resistance was restored in all the src-transfected cell lines while the vector-only controls (2CV or pBABE-1) showed little or no change in oxaliplatin response. This suggests that c-Src is able to prime cells to oxaliplatin-induced apoptosis independently of its kinase domain. This priming for oxaliplatin induced apoptosis is inhibited by CGP77675 by an unclear mechanism that may involve a Src-like tyrosine kinase.

References

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Inhibition of MEK/ERK and PI3K/Akt pathways blocks apoptosis suppression signaling of FLT3/ITD

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Two kinds of FLT3 mutations have recently been detected in patients with acute myeloid leukemia (AML): (1) internal tandem duplication (inserted repeat spanning from as few as 7 to more than 30 amino acids) the juxtamembrane domain in about 20% AML patients and (2) mutations in inactivation loop (generally D835Y) in about 7% of AML patients. Patients harboring FLT3/ITD mutations have relative poor prognosis, especially when the other allele is mutated and/or lost. Both FLT3/ITD and FLT3/D835 mutations result in constitutive autophosphorylation of the receptor. Constitutive activation of FLT3 results in factor independent survival and growth in the transfected 32D and BA/F3 cell lines. Syngeneic mice injected of transfected cells develop myeloproliferative disease. Similarly, transplantation of FLT3/ITDs transfected bone marrow cells causes myeloproliferative disease in another mouse model. These data suggest that FLT3/ITDs inhibit apoptosis, promote proliferation and survival, and lead to leukemogenesis. Normal hematopoietic cells depend on the growth factor for survival and proliferation, whereas leukemic cell lines and primary leukemic cells often become partially or completely growth factor-independent. IL-3 dependent survival in BA/F3 cells is mediated by MEK/ERK pathway and protein kinase A (PKA) pathway. How BA/F3 cells transfected with FLT3/ITD or FLT3/D835 mutation become IL-3 independent is still largely unknown. To understand the mechanism of conversion of FLT3/ITD transfected BA/F3 cells towards IL-3-independence, we investigated (1) the activation of components in PI3K/Akt and MEK/ERK pathways by FLT3/ITD and (2) the effects of the inhibition of FLT3/ITD, PI3 Kinase or MEK on cell survival and proliferation. Our results: (1) FLT3/ITD constitutively activate PI3K/Akt and MEK/ERK pathways; (2) inhibition of FLT3/ITD autophosphorylation eliminated the activation of PI3K/Akt and MEK/ERK pathways and induced rapid apoptosis in most cells; (3) inhibition of PI3 Kinase block cell proliferation but only showed weak effect on apoptosis; (4) inhibition of MEK block proliferation and induced apoptosis; (5) combined inhibition of both PI3 Kinase and MEK had the same strong effects on apoptosis as inhibition of FLT3/ITD. Our results suggested that the mechanism of FLT3/ITD to suppress apoptosis and maintain survival involving both PI3-Kinase/ Akt and MEK/ERK pathways.

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Hypoxia Increases potency of the proteasome inhibitor VELCADETM (bortezomib) for injection: potential for a hypoxic cell cytotoxin in solid tumors

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The 26S proteasome is a ubiquitous enzyme that plays an essential role in the regulation of cellular protein degradation, gene expression and cell cycle transition. In cancer cells these events can be deregulated and it has been hypothesized that antagonism of proteasome function might provide therapeutic benefit. VELCADE, (formerly known as PS-341), a potent and selective inhibitor of the proteasome, induced apoptosis in a variety of human tumor cells and in pre-clinical cancer animal models. In Phase I clinical trials, VELCADE demonstrated promising anti-tumor activity in multiple cancer types and is now in Phase II and III trials. To understand the potential contribution of proteasome function to the observed biological responses to VELCADE treatment, we adapted multiple myeloma cells to proliferate in high concentrations of the drug and generated stable cellular clones with greater than 500-fold resistance. The parental and VELCADE-

adapted myeloma cell lines were subsequently grown as xenografts in mice to evaluate whether the *in vitro* adaptation translates into *in vivo* resistance. Surprisingly, VELCADE showed similar efficacy against tumors generated from both cell lines, suggesting that the tumor microenvironment might influence the efficacy of VELCADE. One parameter evaluated was the potential contribution of tumor hypoxia, a feature of rapidly growing solid tumors which is thought to contribute to the clinical failure of many chemotherapeutics. We found the adapted cells to no longer be resistant to VELCADE-induced apoptosis in hypoxia. Importantly, the parental myeloma cells as well as human lung and colon tumor cells were more sensitive to VELCADE-induced apoptosis under similar hypoxic conditions. These effects were specific for VELCADE, as other proteasome inhibitors and several standard chemotherapeutic agents did not demonstrate increased potency under hypoxic conditions. Since tumor cells often undergo cell cycle arrest under hypoxia, we examined whether VELCADE could induce apoptosis in non-cycling cells. Unlike other chemotherapeutics, VELCADE induced apoptosis equally well in G1-arrested and cycling colon tumor cells. This raises the possibility that VELCADE might work well in combination with hypoxia-inducing anti-angiogenic agents in solid tumors. These findings suggest that VELCADE represents a novel class of cancer therapeutics that offers significant advantages over other therapeutic modalities for the treatment of solid tumors.

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Determinants of AKT-dependent resistance to postmitochondrial apoptosis induction

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In the present study, we examined the molecular mechanisms underlying the chemoresistance of human cancer cells against the topoisomerase II inhibitor etoposide. Using pharmacological inhibitors and after adenoviral gene transfer of AKT signaling mutants, we found that inhibition of AKT activity rapidly and effectively sensitized cancer cells to etoposide-induced apoptosis. To identify the molecular targets of AKT-dependent chemoresistance, we first examined the release of mitochondrial apoptotic complementation factors after etoposide treatment or following AKT inhibition. By using cell fractionation techniques and immunofluorescence analysis, we measured the release of mitochondrial cytochrome c in cancer cell lines including LNCaP prostate carcinoma cells and observed that etoposide treatment alone caused mitochondrial damage independently of AKT inhibition. The subsequent activation of caspase-3/7 and apoptosis, however, was not induced efficiently unless AKT activity was also suppressed. Thus, we concluded that at least in some cancer cell lines, AKT inhibition was required to facilitate caspase activation by apoptogenic mitochondrial factors. To determine the mechanism underlying the AKT-dependent postmitochondrial resistance, we then examined whether downstream caspases were inhibited by direct AKT-dependent phosphorylation. By using transient transfection assays and *in vitro* reconstitution experiments using AKT-specific phosphorylation site mutants, we determined that the direct inhibition of caspases by AKT did not play a significant role in determining the AKT-dependent outcome after etoposide treatment. We also found that the AKT-dependent sensitization to apoptosis induction, even though it required ongoing macromolecular synthesis, did not depend on p53-dependent transcriptional activity. Supported in part by DAMD17-99-1-9153 and DAMD17-00-1-0214, and by the Speaker's Fund for Biomedical Research.

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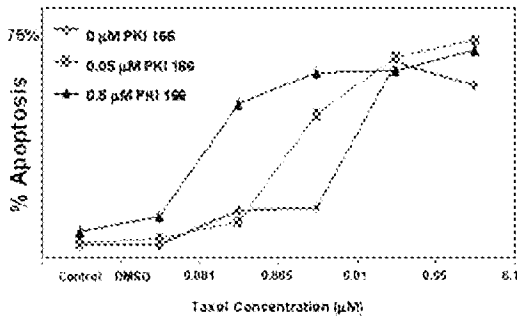
Synergistic therapy of head and neck cancer with EGFR blockade and paclitaxel is mediated through abrogation of PI3kinase/AKT signaling

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Purpose: Survival for patients with oral cancer has not improved over the past 25 years and new approaches for treatment are needed. Targeted molecular therapy against epidermal growth factor receptor (EGFR) has shown promise as an adjuvant therapy in preliminary studies in several solid tumors, including head and neck cancer. The objective of this study is to determine the efficacy of paclitaxel and PK1166, a novel inhibitor of EGFR, in oral cavity cancer.

Experimental Design and Results: Using propidium iodide assay, JMAR human oral cancer cells were evaluated for the induction of apoptosis when treated with paclitaxel [0.001 to 0.1 mM] in the absence or presence of

PKI166 [0, 0.05, 0.5 mM]. The addition of 0.5mM PKI166 increased tumor cell death in a synergistic fashion, shifting the amount of paclitaxel needed to induce apoptosis in 50% of cells from 0.1 to 0.001 mM.



Graph 1. Synergistic induction of apoptosis.

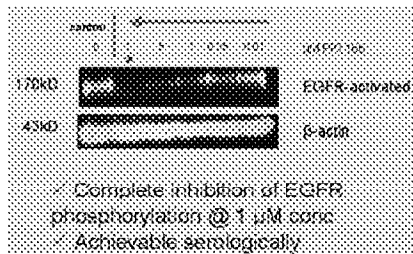


Image 1. PKI166 inhibits EGFR signalling.

Next, JMAR were pretreated for 1 hour and then stimulated with EGF. As measured by Western immunoblotting, EGF-R-specific tyrosine kinase autophosphorylation was inhibited by PKI166 in a dose-dependent fashion and at all doses tested (0.01 to 1 mM). PKI166 also inhibited phosphorylation of AKT (ser473) for in a dose-dependent manner.

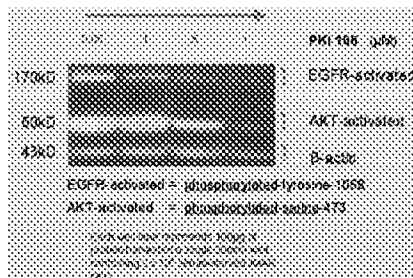
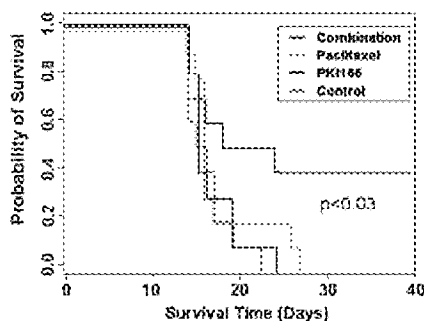


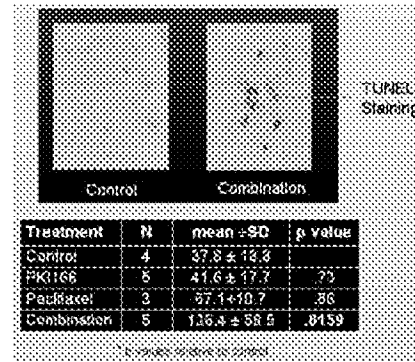
Image 2

Finally, these findings were confirmed using an orthotopic model of oral cancer. JMAR oral cancer cells were implanted in the tongues of nude mice. After lingual tumors developed, 40 mice were randomized into four groups. The animals were treated with 1) oral PKI166 [100mg/kg]; 2) intraperitoneal paclitaxel [2 mg/kg]; 3) PKI166 and paclitaxel; or 4) placebo and followed for survival. Mice treated with PKI166/paclitaxel demonstrated a significant increase in survival ($p=0.028$).



After necropsy, all tongue tumors were evaluated for apoptosis by TUNEL assay and examined under fluorescence microscopy. Tumor specimens

from mice treated with paclitaxel and PKI166 revealed a greater apoptotic fraction of tumor cells than mice receiving placebo or single-drug therapy (136.4 vs. 37.8; $p=0.016$).



Conclusions: Combination therapy with paclitaxel and PKI166 leads to prolonged survival in an orthotopic preclinical model of tongue cancer. This targeted molecular therapy increases programmed cell death of oral cancer *in vitro* and *in vivo* and may be mediated through abrogation of PI3kinase/AKT signaling.

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β2-Microglobulin induces apoptosis in CCRF-HSB-2 leukemia cells by molecular mechanisms different than doxorubicin- and Taxol-induced apoptosis mechanisms

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We have previously been shown that β2-microglobulin (β2m) triggers apoptosis in cancer cells, but the molecular mechanism of β2m-induced apoptosis is not clear. In this study, we investigated the mechanism of β2m-induced apoptosis in the CCRF-HSB-2 human lymphoblastic leukemia cell line and compared it to the apoptotic effects of doxorubicin (DOX) and Taxol in these cells. We found that exogenous β2m induces significant apoptosis in this cell line. β2m treatment induced the release of cytochrome c from the mitochondria, but no change in mitochondrial membrane potential was observed during apoptosis, suggesting that cytochrome c may be released through a mechanism independent of mitochondrial permeability transition (MPT) pore formation. Moreover, the β2m-induced release of cytochrome c from the mitochondria in these cells was caspase-independent, since Z-VAD-fmk, a general inhibitor of caspases, did not block the release of these factors. However, Z-VAD-fmk treatment significantly blocked β2m-induced apoptosis while Western blot analysis revealed that caspases-1, -2, -3, -6, -8, -9, and -10 are not activated during β2m-induced apoptosis in these cells. These results indicate that a caspase-dependent mechanism independent of these caspases is involved in β2m-induced apoptosis. Interestingly, both DOX and Taxol induced apoptosis and activated caspases-6 and -8, indicating that these drugs cause apoptosis by a different mechanism than β2m. Moreover, β2m significantly enhanced the production of reactive oxygen species (ROS) during 12-48 hr treatment, and β2m-induced apoptosis was almost totally inhibited in cells pre-treated with the antioxidant N-acetylcysteine (NAC), providing evidence that β2m-induced apoptosis in these cells is ROS-dependent. Therefore, these results reveal that β2m-induced apoptosis in CCRF-HSB-2 cells occurs through an unknown caspase-dependent, ROS-dependent mechanism(s) which is associated with cytochrome c release from mitochondria. Based on their different apoptosis mechanisms, combination use of β2m and these anticancer agents may provide a more effective therapeutic strategy to eliminate cancer cells. (Supported by grant CA080734 from NCI).